

Multiple Sequence Alignments

A Brief Introduction

EMBL-Australia Masterclass on Protein
Sequence Analysis

Sydney, Australia
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EMBL Heidelberg

Session Goal

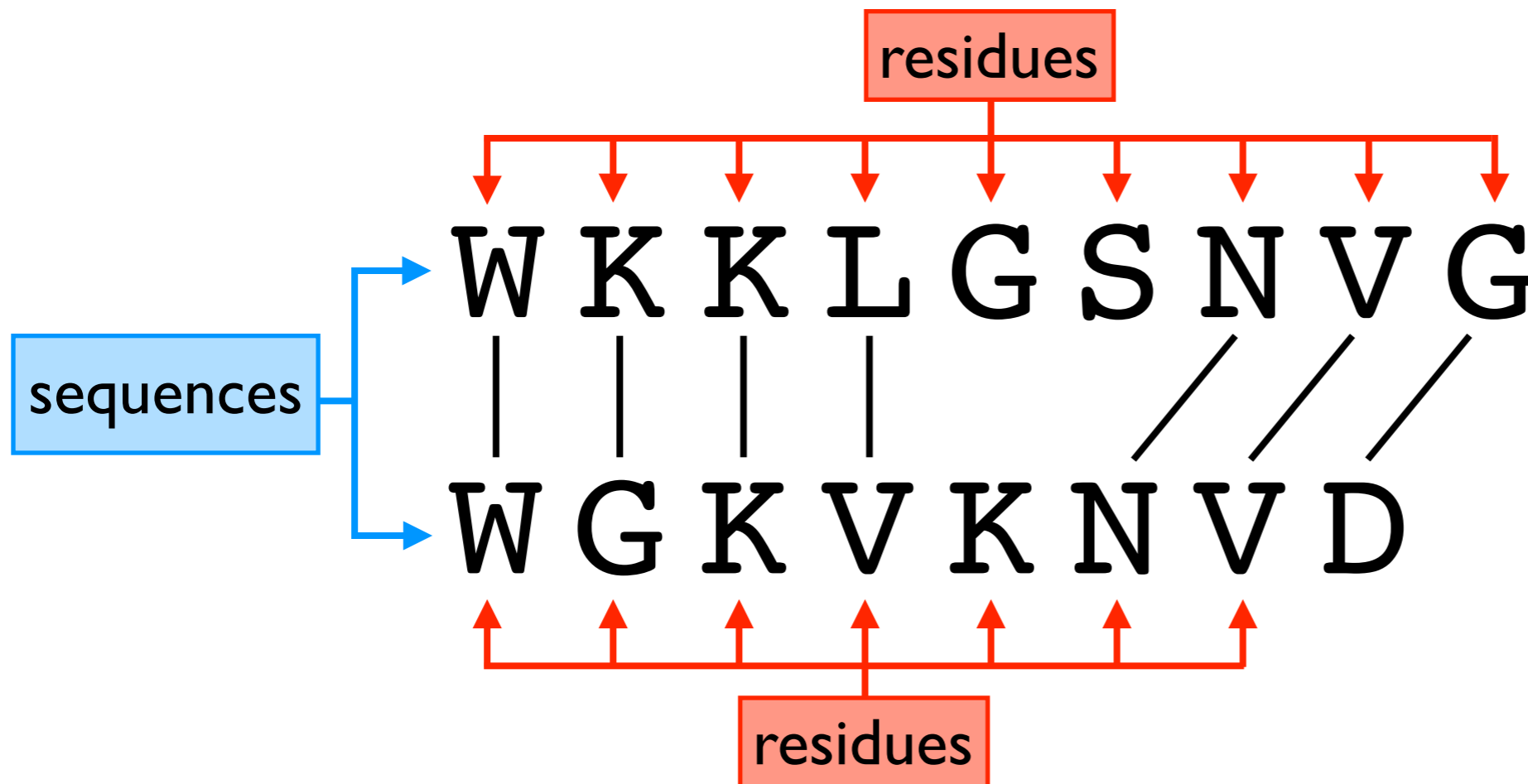
After attending today's session, we hope you will be better able to:

- build higher-quality/more appropriate MSAs for use in your own research/applications
- critically assess the quality of MSAs built by yourself and others

Why a Session on MSAs?

- Required for the development of almost all sequence analysis bioinformatics/tools
- MSAs take practice to interpret (and build) well
- Quality of downstream analysis/tools depends on quality of MSA

"Anatomy" of a Sequence Alignment



Residues:

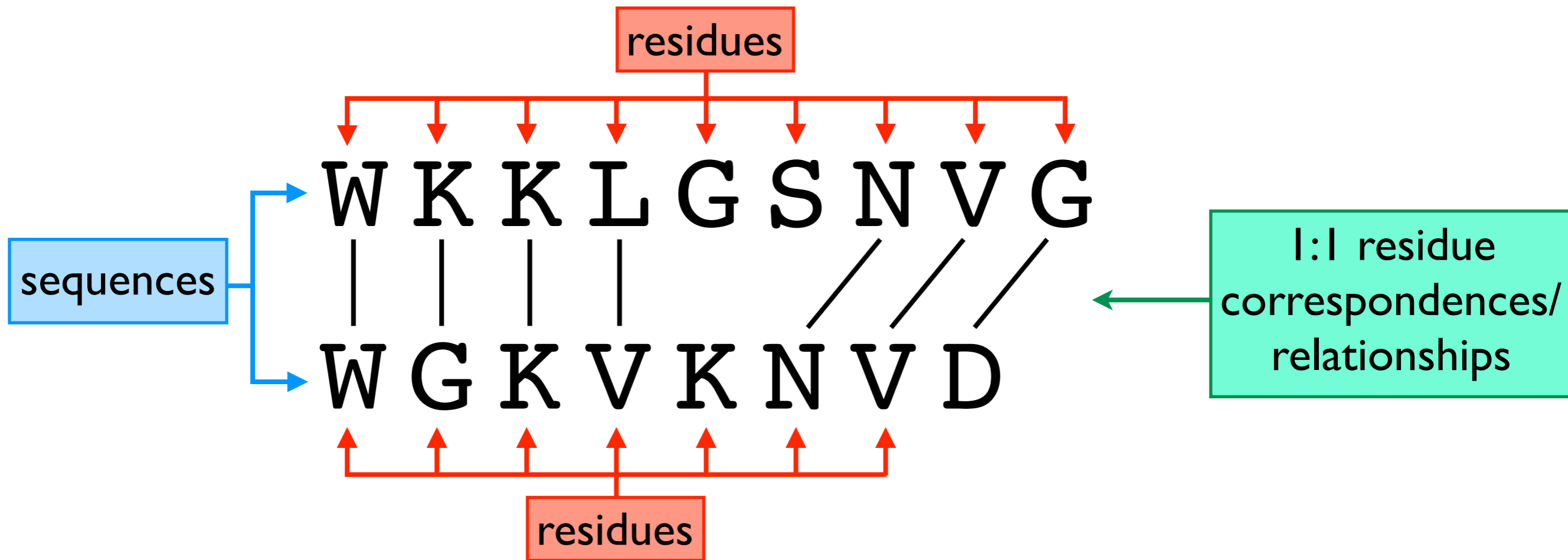
Monomers within a polymer (polypeptide or polynucleotide) chain

Sequences:

List of residues in a polymer chain...

...listed in the same order they occur within the polymer

"Anatomy" of a Sequence Alignment

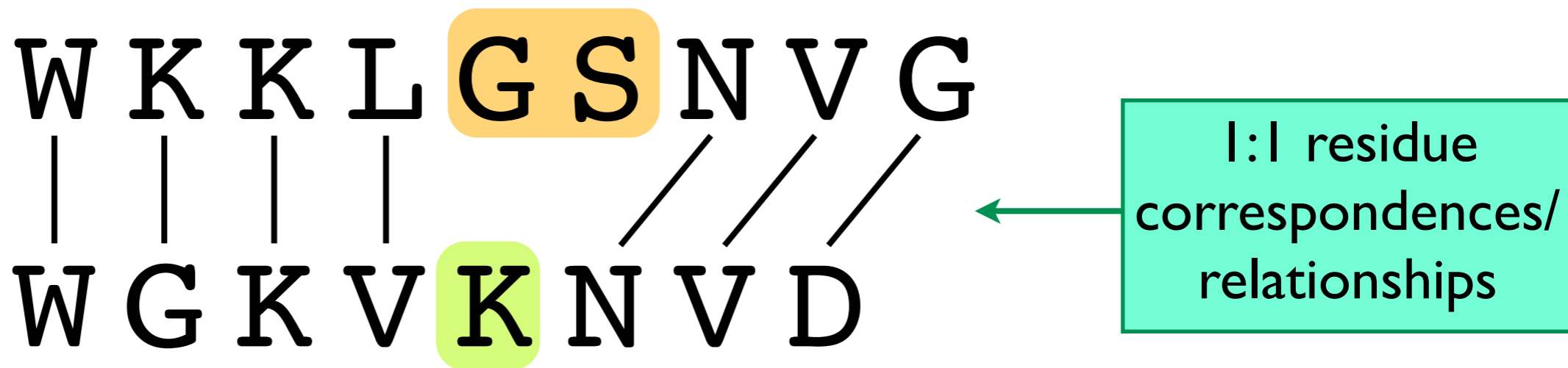


1:1 residue correspondences/relationships

Correspondences between

- a single residue in one sequence and
- a single residue in another sequence

"Anatomy" of a Sequence Alignment



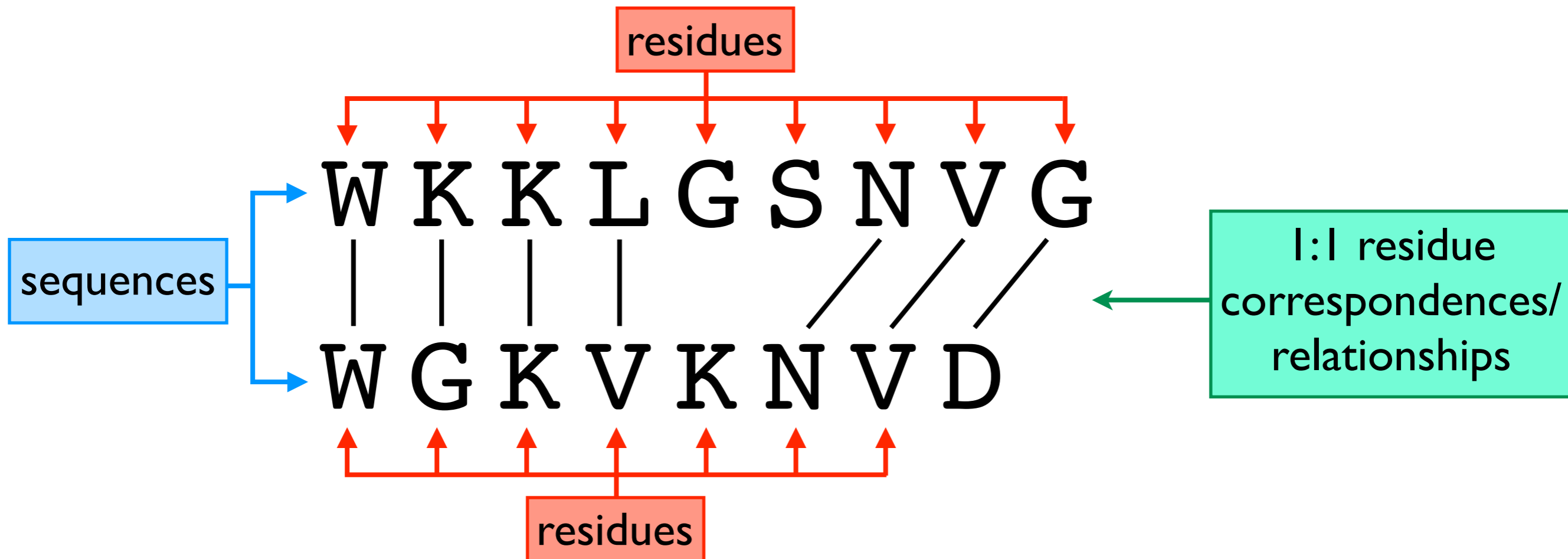
Residue has no equivalent in the top sequence

i.e. no residue in the top sequence has a 1:1 relationship with this residue

Could *perhaps* say there is a "1:2" relationship between **this residue** and **these residues**

However, alignments focus on 1:1 relationships

"Anatomy" of a Sequence Alignment

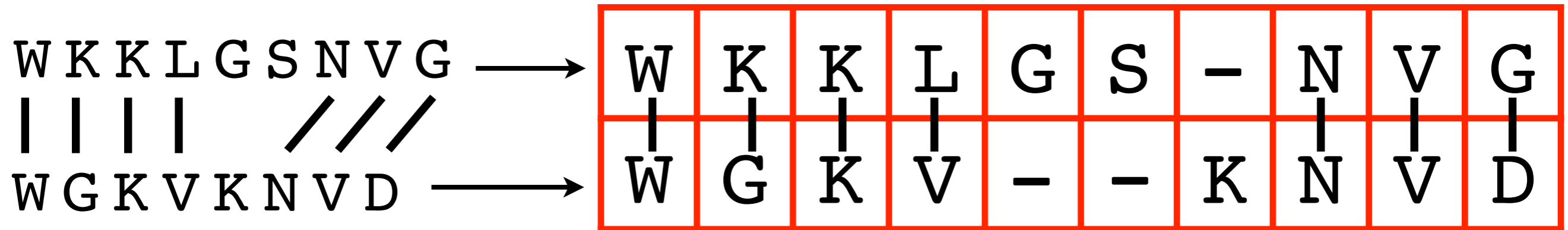


Sequence alignment

A comparison of the residues in two or more sequences...

...describing 1:1 correspondences/relationships/equivalences between residues in different sequences

Sequence Alignment Within a Grid



Often represented using a **grid/matrix**:

One sequence per row

Residues in the same column are 'equivalent'

Gap characters (usually "-") indicate that the sequence contains no residues 'equivalent' to other residues in that column

Alternative Interpretations of MSAs (Evolutionary and Structural)

“Equivalence”/similarity of residues

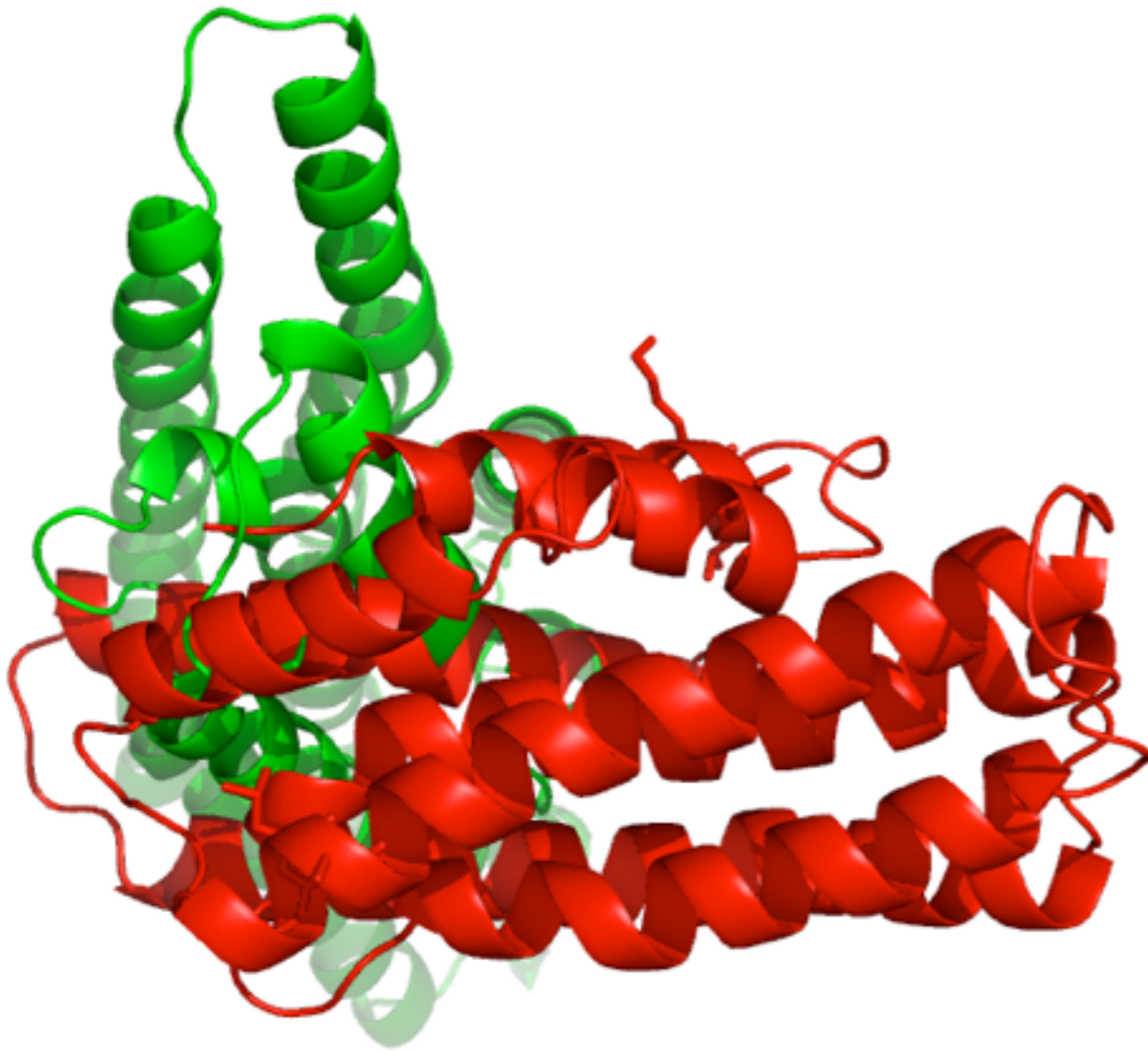
Residues in the same column either:

- Structurally equivalent/similar
- Evolutionary equivalent/related/homologous

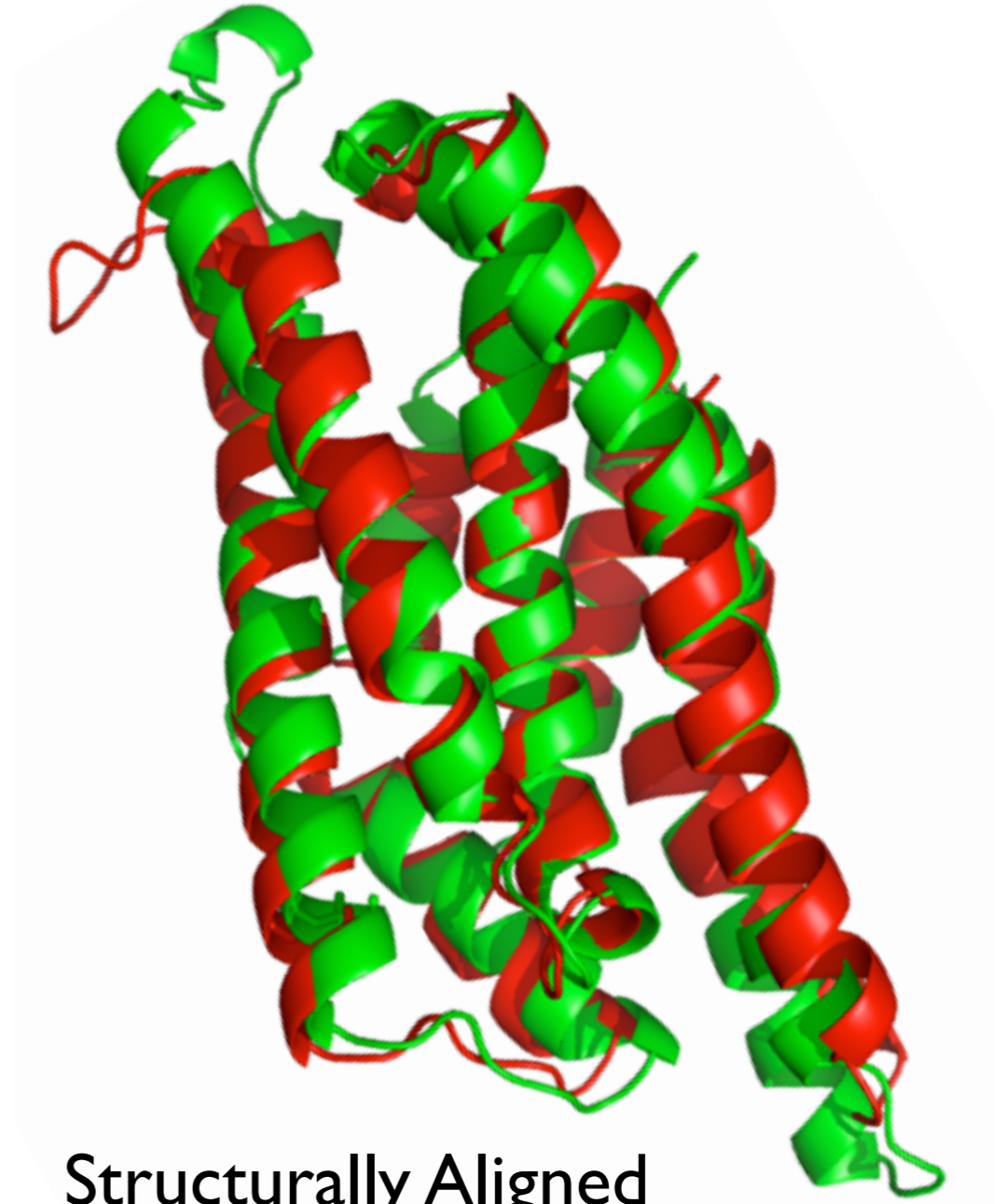
Different applications assume different types of equivalence

Different types of similarity not necessarily equivalent

Structural Similarity



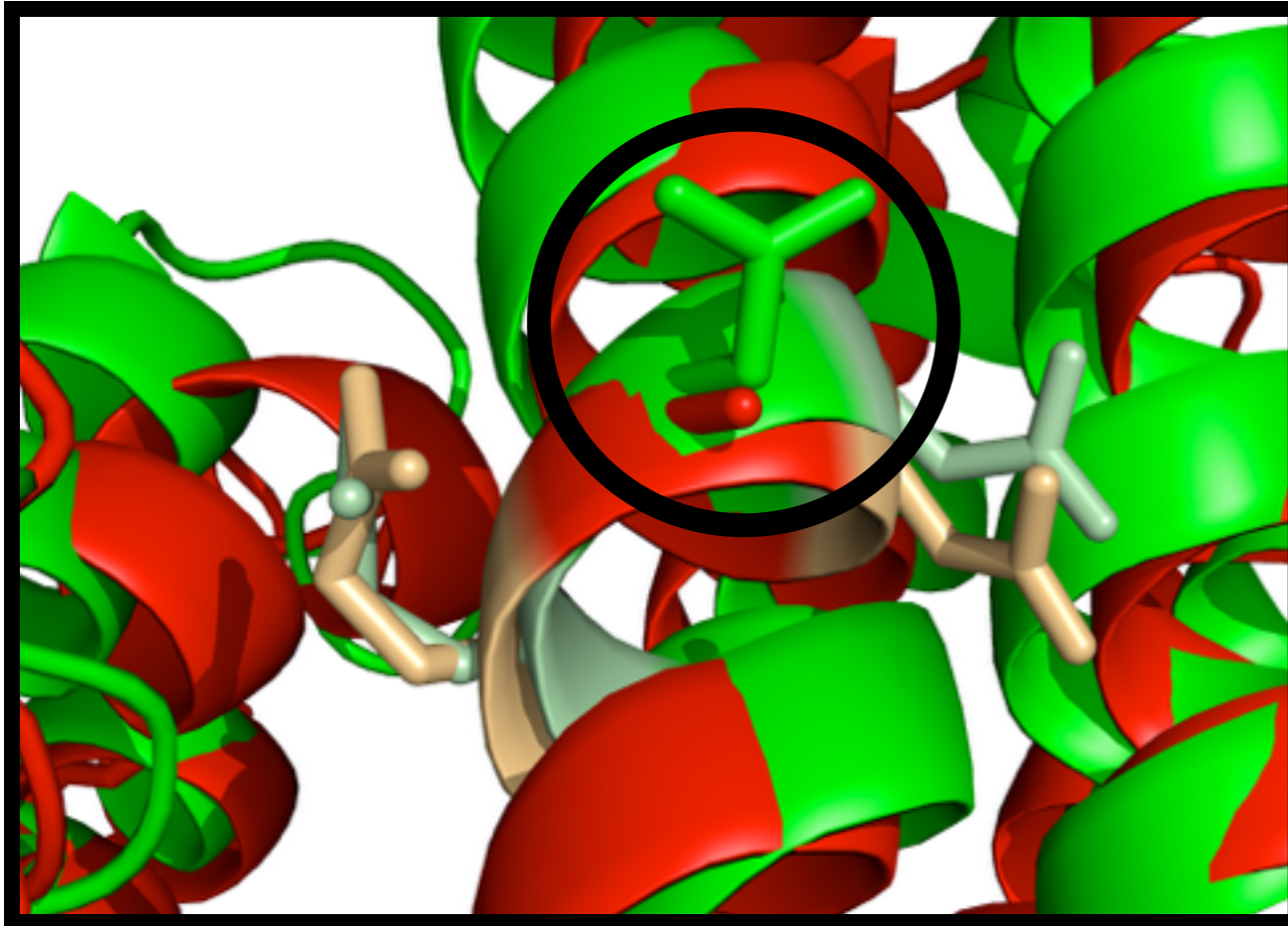
Unaligned



Structurally Aligned

Bacterial toxins **Iji6** and **Ii5p**

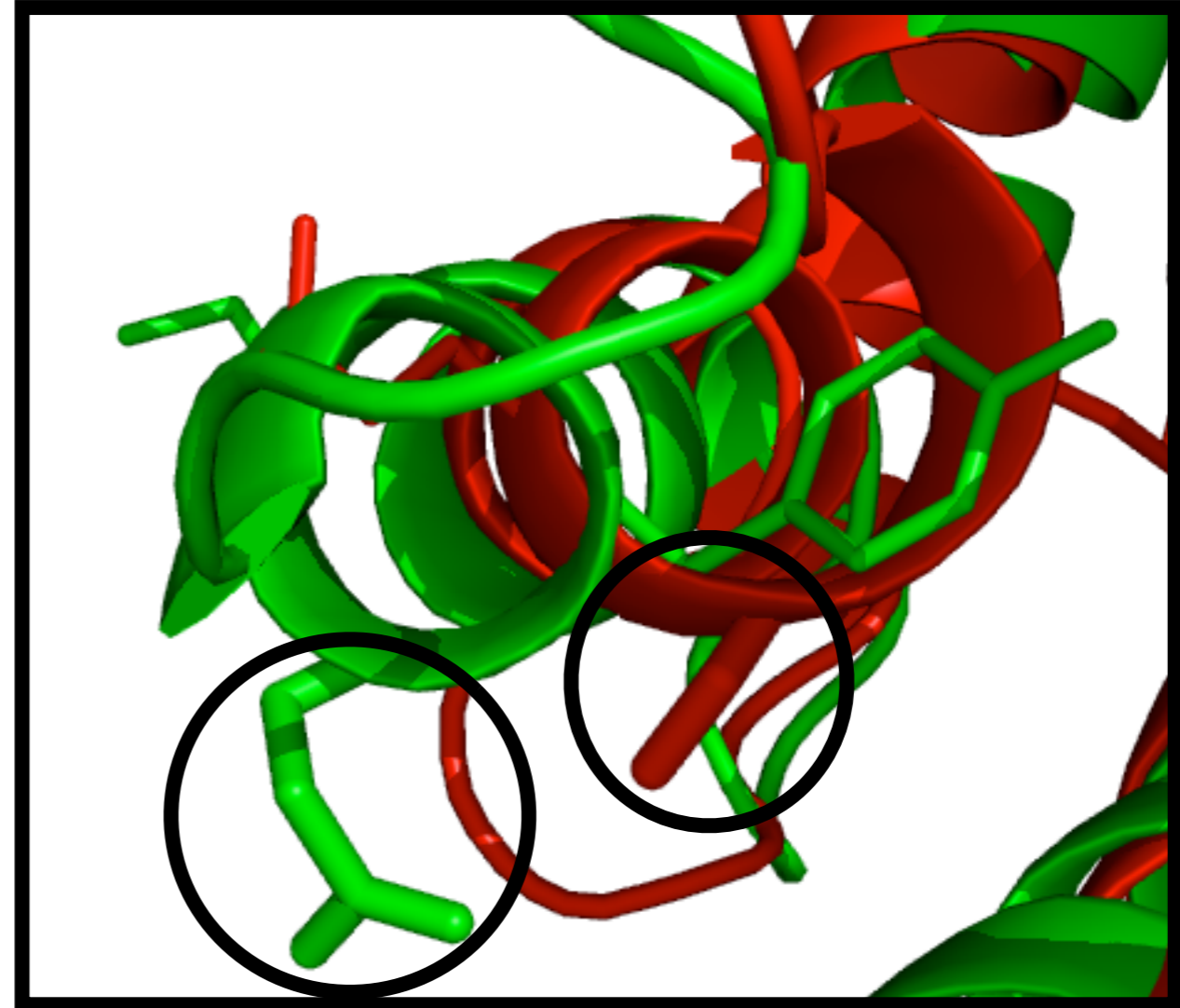
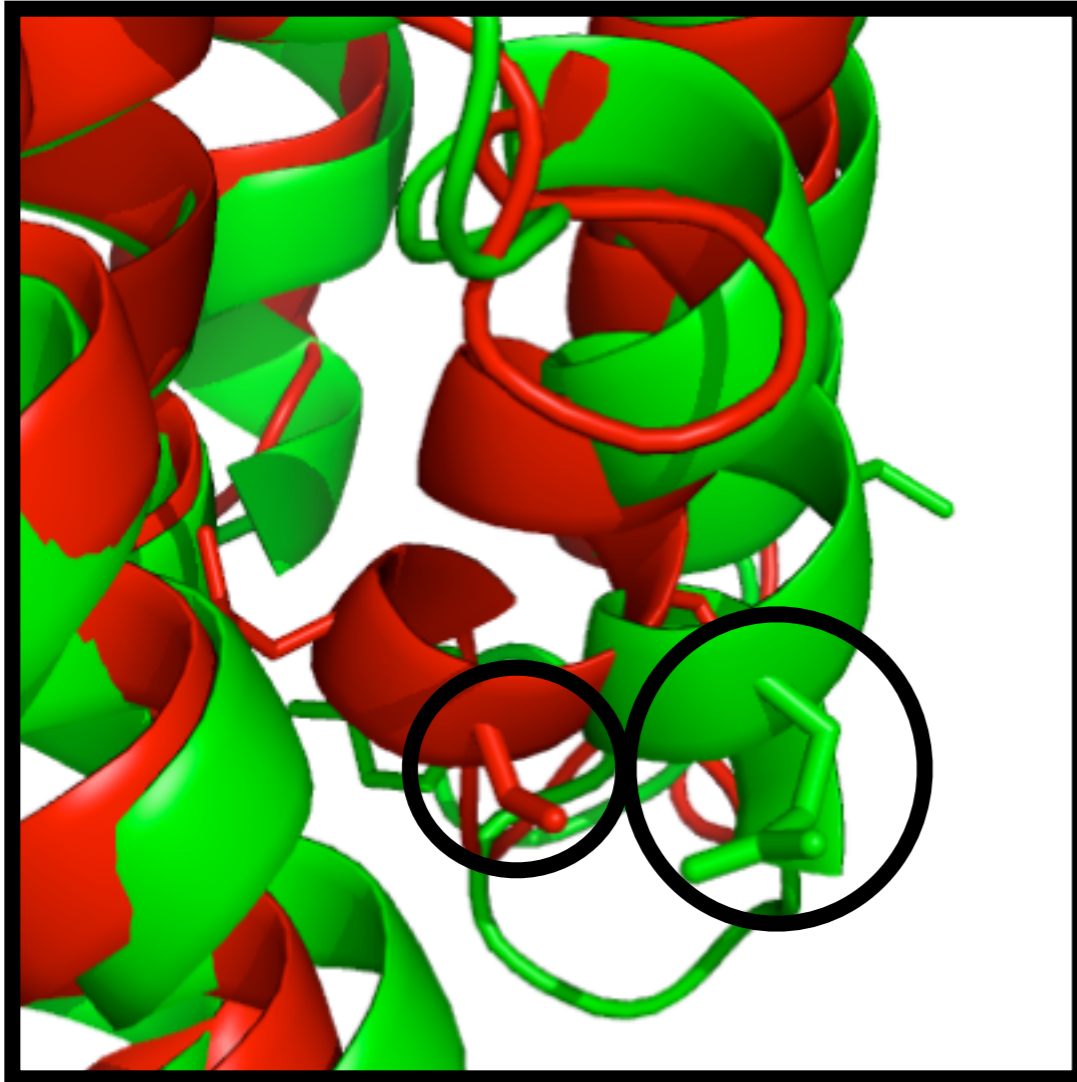
Structural Similarity



68 ELIGLQANIREFNOQVDNF
11111111111111111111
70 ELQGLQNNFEDYVNALNSW

Residues with a similar structural context may lie almost on top of each other within a structural alignment. Clearly, the dark green and red side chains have more similar structural contexts than they do with the adjacent light-coloured side chains

Structural Similarity



```
Chain 1: 16 KVGSLIGKR---ILSELWGIIFPSGST  
          111111111 11111111111 111  
Chain 2: 16 VVGVPFAGALTSFYQSFLNTIWP-SDA
```

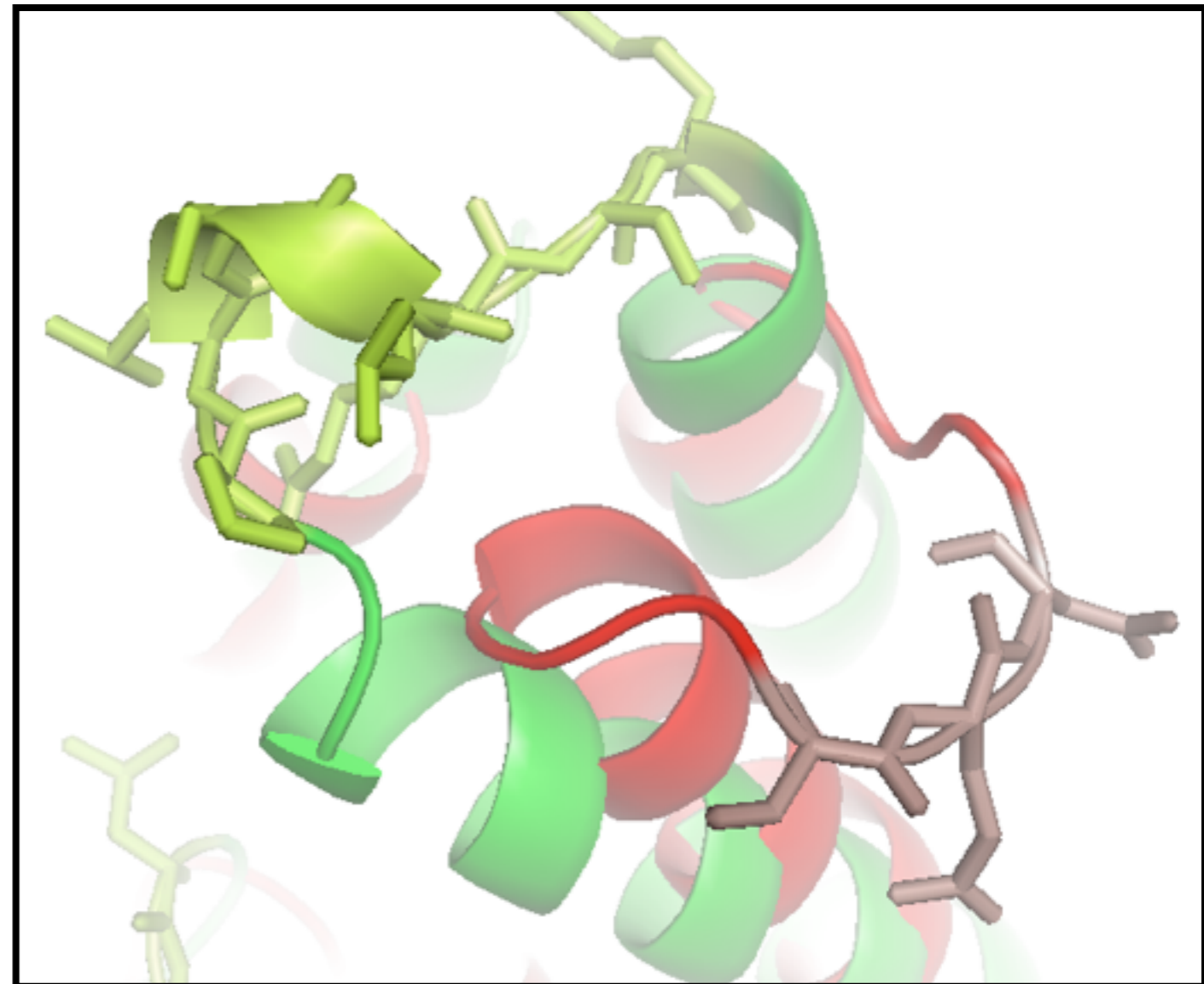
Structural equivalence

Some regions of the structures do not have structurally equivalent residues in the other structure

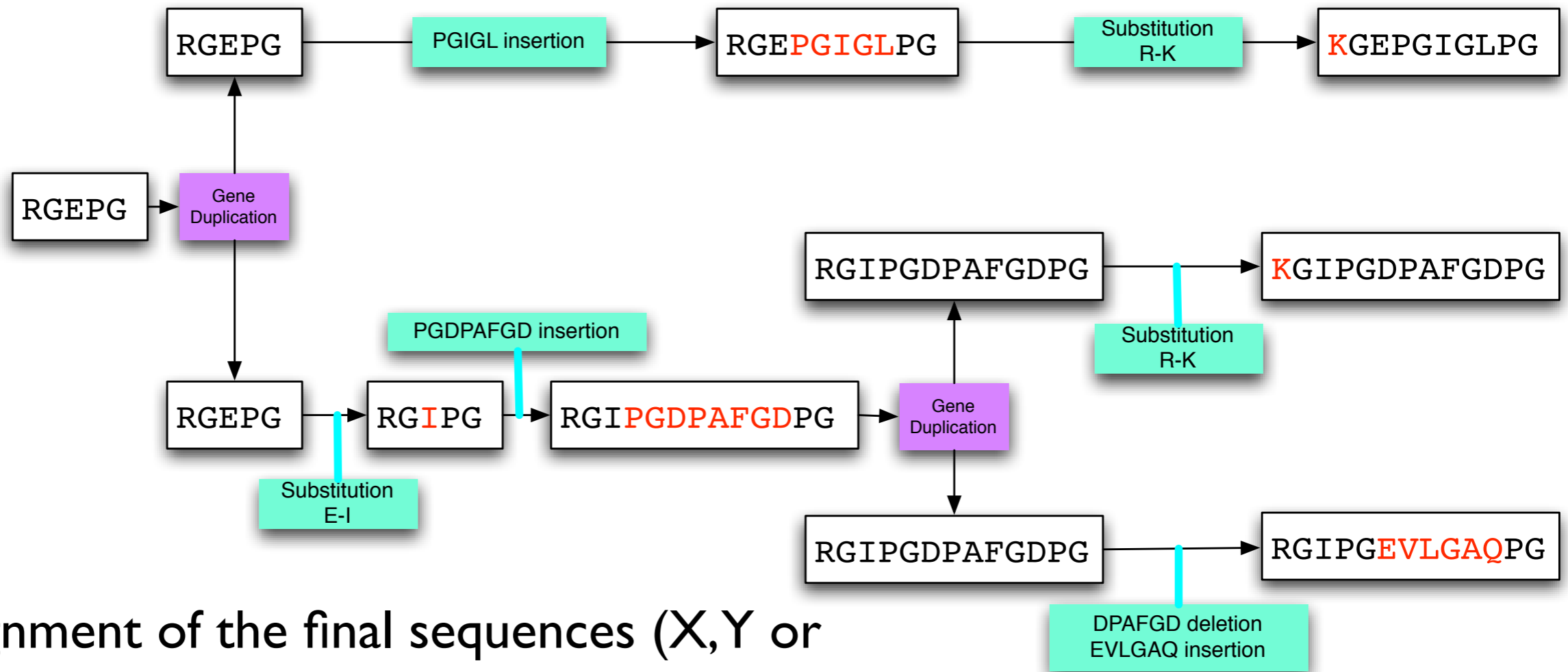
Alignment gaps are a sure sign of such residues

Placing such residues in the same column as residues from other sequences is a **misalignment** - to be avoided!

```
1i5p:  DNFLNPTQN-----PVPLSITSSVN
      111111          111111111111
1ji6:  NSWKKTPLSLRSKRSQDRIRELFS
```



Quiz - Evolutionary Interpretation of Alignments



Which alignment of the final sequences (X, Y or Z) only places residues in the same column if they are related by substitution events?

X

KGEPG---IGLPG
 KGIPGDPAFGDPG
 RGIPGEVLGAQPG

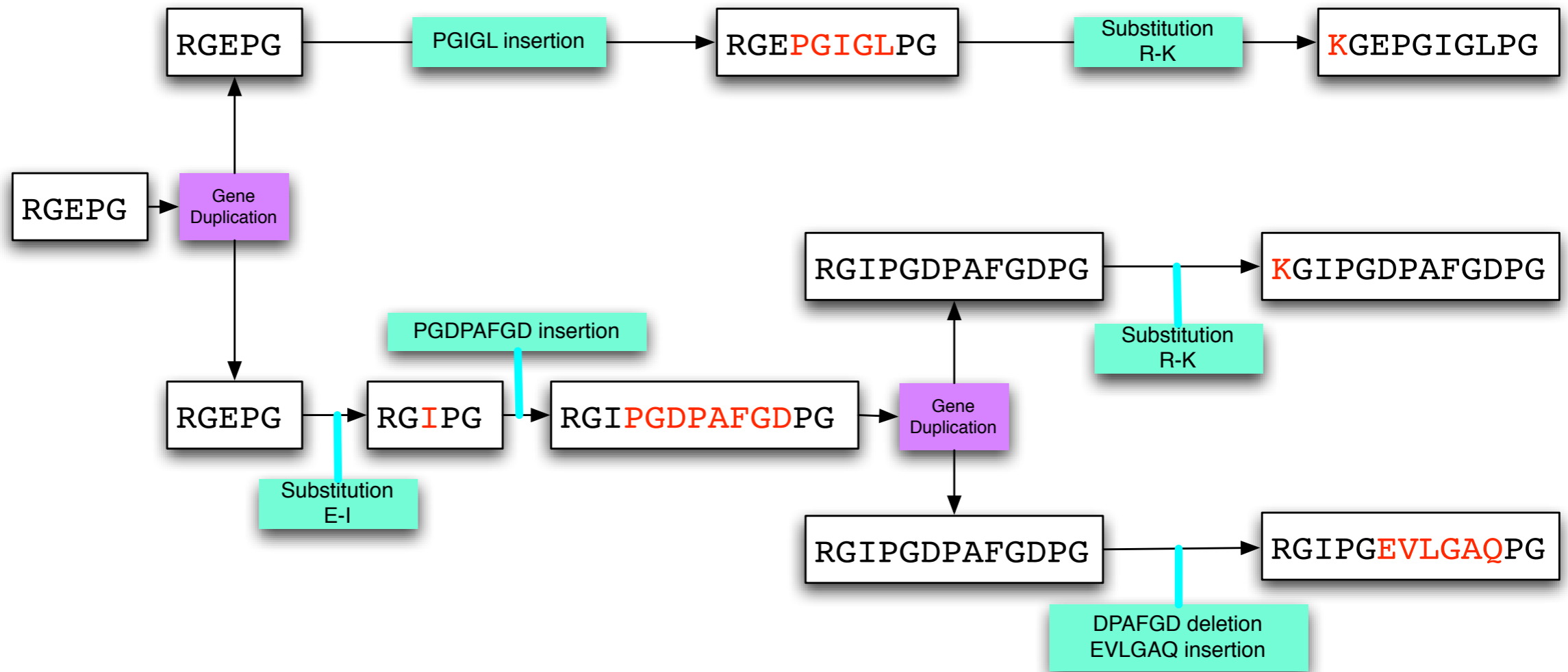
Y

KGEPG-----IGL-----PG
 KGIPG-----DPAFGDPG
 RGIPGEVLGAQ-----PG

Z

KGE-----PGIGL-----PG
 KGIPG-----DPAFGDPG
 RGIPGEVLGAQ-----PG

Quiz - Evolutionary Interpretation of Alignments



"True" alignment given history described above

```

KGE-----PGIGL-----PG
KGIPG-----DPAFGDPG
RGIPGEVLGAQ-----PG
    
```

PRANK

```

RGIPGEVLGAQPG
KGIPGDPAFGDPG
---KGEPGIGLPG
    
```

Quiz - Evolutionary Interpretation of Alignments

CLUSTALX

```
K---GEPGIGLPG
KGIPGDPAFGDPG
RGIPGEVLGAQPG
```

MAFFT

```
KGEPG---IGLPG
KGIPGDPAFGDPG
RGIPGEVLGAQPG
```

PRANK

```
RGIPGEVLGAQPG
KGIPGDPAFGDPG
---KGEPGIGLPG
```

Different automatic MSA software gives different results

All are different from the "true" alignment (assuming the scenario of transformation on the previous slide is true)...

... because that scenario is very unlikely under the models of evolutionary transformation incorporated within these tools

X

```
KGEPG---IGLPG
KGIPGDPAFGDPG
RGIPGEVLGAQPG
```

Y

```
KGEPG-----IGL-----PG
KGIPG-----DPAFGDPG
RGIPGEVLGAQ-----PG
```

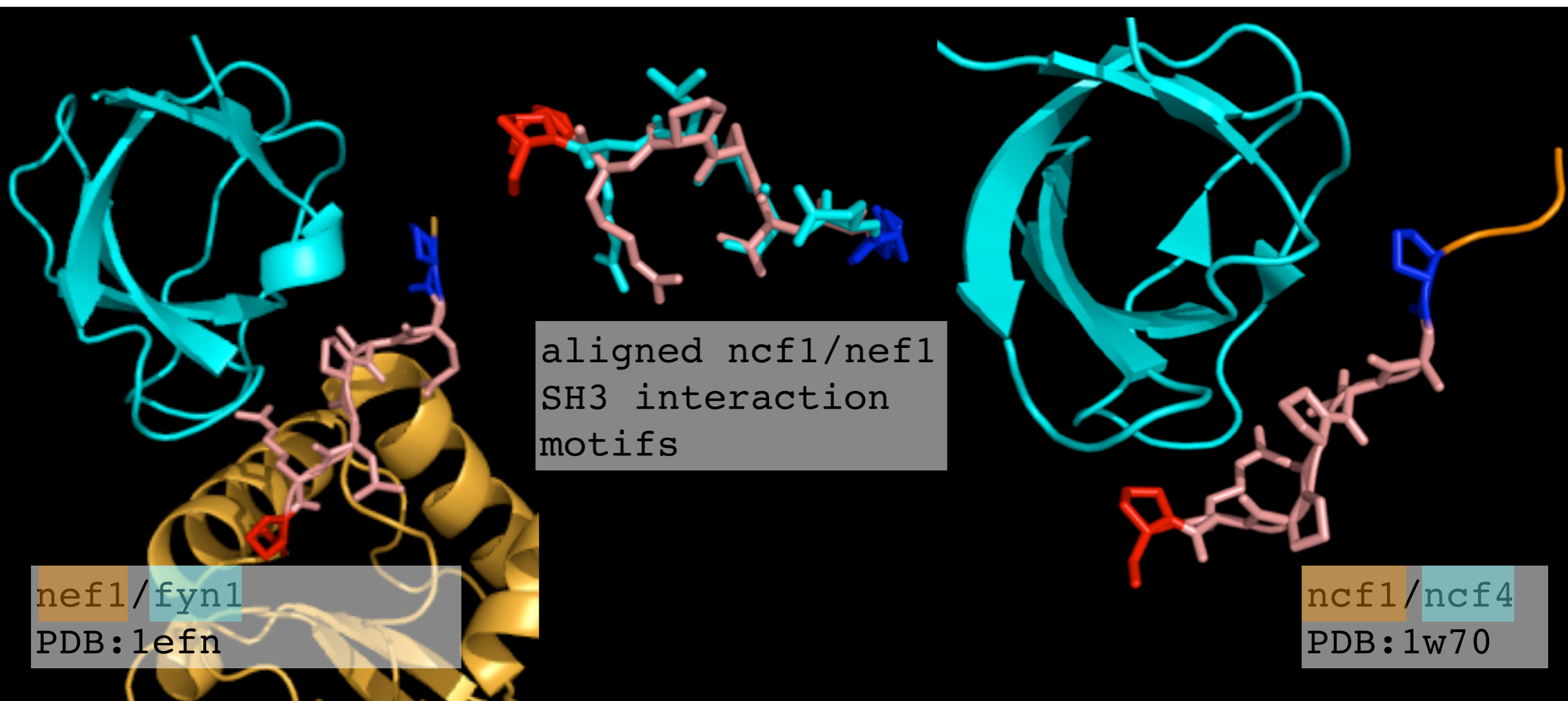
Z

```
KGE-----PGIGL-----PG
KGIPG-----DPAFGDPG
RGIPGEVLGAQ-----PG
```

Non-Equivalence of Evolutionary and Structural Alignments

Structural equivalence without evolutionary equivalence

Structural alignment of SH3-interaction motifs from nef and ncf1

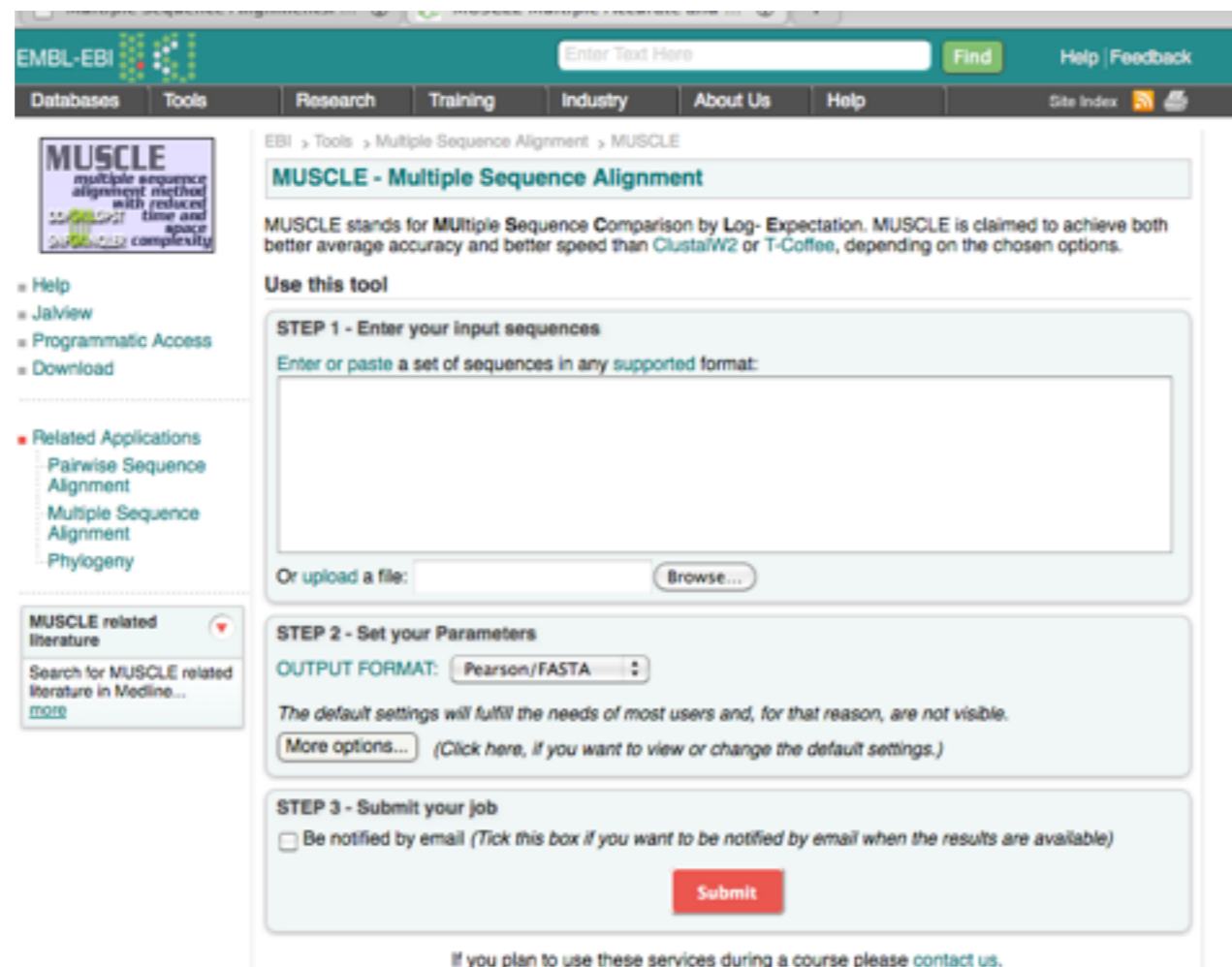


Building MSAs

Build an Automatic MSA

Load sequences into JalView, and with a few clicks you can automatically align a set of sequences

Or run an MSA tool at EBI

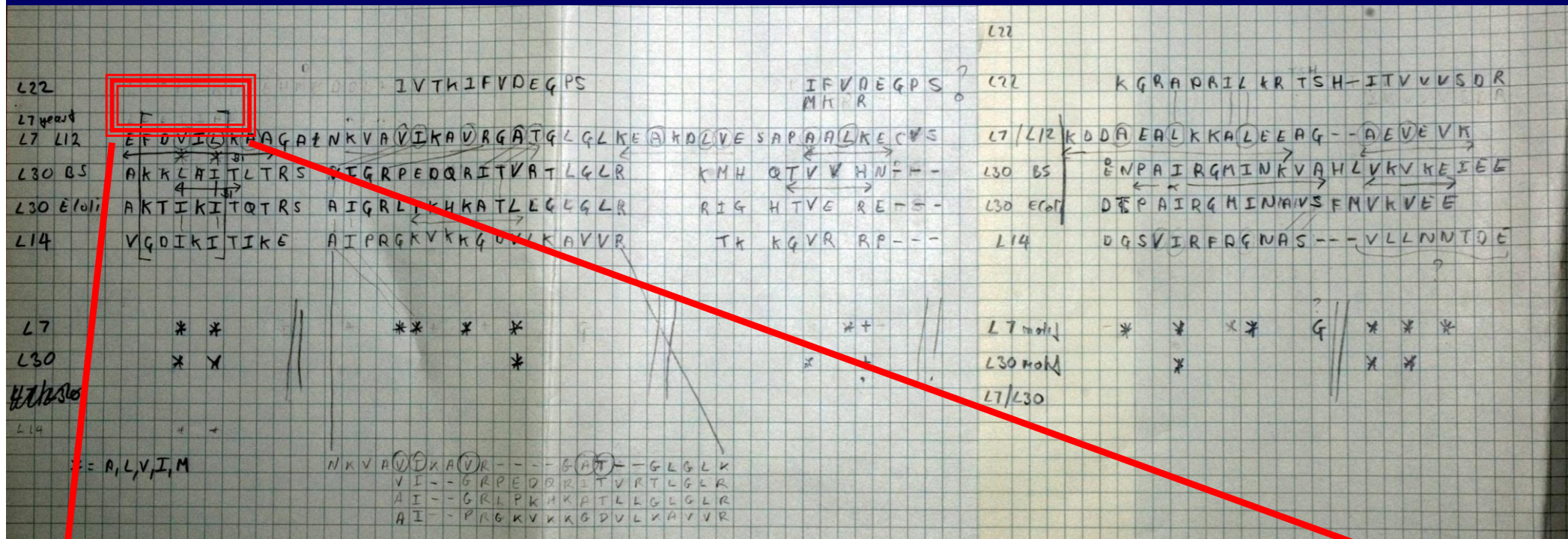


The screenshot shows the EBI MUSCLE web interface. At the top, there is a navigation bar with 'EMBL-EBI' logo, a search box, and links for 'Databases', 'Tools', 'Research', 'Training', 'Industry', 'About Us', and 'Help'. Below the navigation bar, the main content area is titled 'MUSCLE - Multiple Sequence Alignment'. It includes a brief description of the tool, a 'Use this tool' section with three steps: 'STEP 1 - Enter your input sequences' (with a text input field and a 'Browse...' button), 'STEP 2 - Set your Parameters' (with an 'OUTPUT FORMAT' dropdown set to 'Pearson/FASTA' and a 'More options...' link), and 'STEP 3 - Submit your job' (with a checkbox for email notifications and a 'Submit' button). A sidebar on the left contains links for 'Help', 'JalView', 'Programmatic Access', 'Download', 'Related Applications' (Pairwise Sequence Alignment, Multiple Sequence Alignment, Phylogeny), and 'MUSCLE related literature'.

<http://www.ebi.ac.uk/Tools/msa/muscle/>

Build an MSA "Manually"

Multiple Sequence Alignment and Visualisation (1984/5)



Courtesy of Geoff Barton, Dundee

JaView Demo and Exercises

- Loading sequences
- Changing the way the sequences are displayed
- Manual editing of alignments
- Adding/removing sequences to an alignment
- Exporting sequences/alignments from JaView for use in another application

JalView Demo and Exercises

- a process of pattern-matching/identification
- we prefer alignments where many columns contain few differences/conservative changes
- more divergent sequences are harder to align than more similar sequences
 - for divergent sequences, there are many alternative alignments are similarly good/bad
 - for rather similar sequences, there is usually one/a few alignments we feel are clearly much better than the others
- Longer sequences take longer to align than short ones

JalView Demo and Exercises

- More sequences take longer to align than fewer sequences
- Repeats cause problems
- Different positions evolve differently
- At some level, the problem is "simple"
 - we just have to choose the right place to put the gaps!

another quiz on interpreting MSAs...

Quiz - Numbers of Insertions

```
mouseHemoglobinB1 AVSCLWGKV--NSDEVGGEALGRL
mouseHemoglobinBZ AITSIWDKV--DLEKVGGETLGRRL
mouseHemoglobinE  LINGLWSKV--NVEEVGGEALGRL
humanHemoglobinAZ IIVSMWAKISTQADTIGTETLERL
mouseHemoglobinAZ IIMSMWEKMAAQAEPIGTETLERL
humanHemoglobinG2 TITSLWGKV--NVEDAGGETLGRRL
humanHemoglobinAT LVRALWKKLGSNVGVYTTTEALERT
humanHemoglobinA  NVKAAWGVGAHAGEYGAALERL
humanHemoglobinB  AVTALWGKV--NVDEVGGEALGRL
```

The **minimum** number of insertion events required to account for the section of haemoglobin alignment shown above is?

(a) 2

(b) 1

(c) 0

(d) 3

Quiz - Numbers of Insertions

```
mouseHemoglobinB1 AVSCLWGKV--NSDEVGGEALGRL
mouseHemoglobinBZ AITSIWDKV--DLEKVGGETLGRRL
mouseHemoglobinE  LINGLWSKV--NVEEVGGEALGRL
humanHemoglobinAZ IIVSMWAKISTQADTIGTETLERL
mouseHemoglobinAZ IIMSMWEKMAAQAEPIGTETLERL
humanHemoglobinG2 TITSLWGKV--NVEDAGGETLGRRL
humanHemoglobinAT LVRALWKKLGSNVGVYTTTEALERT
humanHemoglobinA  NVKAAWGVGAHAGEYGAEALERM
humanHemoglobinB  AVTALWGKV--NVDEVGGEALGRL
```

The **minimum** number of insertion events required to account for the section of haemoglobin alignment shown above is?

If all sequences are the same length, we can explain their diversity without inferring **ANY** insertions or deletions

If an alignment contains sequences that are all either length **x** or **y**, then we can explain their diversity by inferring just one insertion or deletion

Quiz - Numbers of Insertions

```
mouseHemoglobinB1 AVSCLWGKV--NSDEVGGEALGRL
mouseHemoglobinBZ AITSIWDKV--DLEKVGGETLGRRL
mouseHemoglobinE  LINGLWSKV--NVEEVGGEALGRL
humanHemoglobinAZ IIVSMWAKISTQADTIGTETLERL
mouseHemoglobinAZ IIMSMWEKMAAQAEPIGTETLERL
humanHemoglobinG2 TITSLWGKV--NVEDAGGETLGRRL
humanHemoglobinAT LVRALWKKLGSNVGVYTTTEALERT
humanHemoglobinA  NVKAAWGVGAHAGEYGAEALERM
humanHemoglobinB  AVTALWGKV--NVDEVGGEALGRL
```

The **minimum** number of insertion events required to account for the section of haemoglobin alignment shown above is?

We can **ALWAYS** explain observed sequence length diversity with:

- 0 insertions (all length variation due to deletion)
- 0 deletions (all length variation due to insertion)
- a combination of insertions and deletions

Perhaps we should instead focus on inferring the **most likely** scenario?

(Although if this is not particularly relevant for our analysis, perhaps we should focus instead on something completely different!)